



Endo- β -Galactosidase

Source

Recombinant from *Bacteroides fragilis*

Catalog Number

E-XBG01

Certification of Analysis Lot Number

512.1B

Contents

1 vial: Endo- β -Galactosidase- 60 μ L (0.9 U)
in 20 mM tris-HCl, pH 7.5

1 vial: Reaction buffer - 400 μ L
250mM Sodium phosphate, pH 5.8

Specific Activity 150 U/mg

Activity 15 U/ml

pH Optimum 5.8

MW 32,000 daltons

Formulation

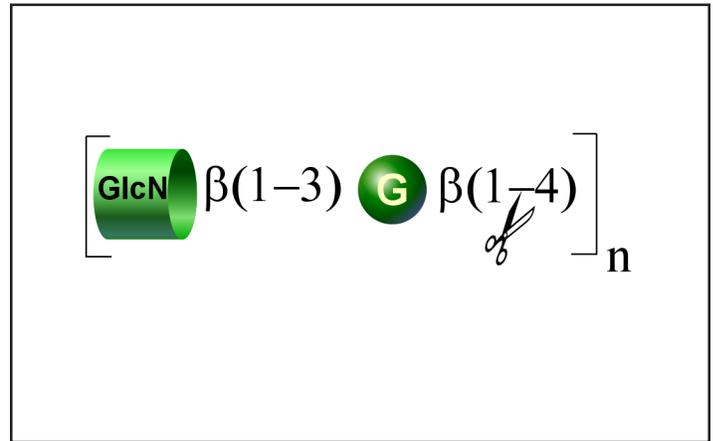
The enzyme is provided as a sterile-filtered solution in 20 mM Tris-HCl, pH 7.5

Storage

Store enzyme at 4°C. Do not freeze.

Stability

Stable at least 12 months when stored properly. Several days exposure to ambient temperatures will not reduce activity. Active at least 5 days under reaction conditions.



Applications

Endo- β -Galactosidase (EC 3.2.1.103) cleaves internal β (1-4) galactose linkages in unbranched, repeating poly-N-acetyllactosamine structures. Sulfated structures such as keratan sulfate are also cleaved. Branching and/or fucosylation of the substrate may decrease or eliminate cleavage.

Endo- β -Galactosidase is useful for identifying and removing poly-N-acetyllactosamine structures on many biologically important glycoconjugates.

Specificity

Internal β (1-4) galactose linkages in unbranched, repeating poly-N-acetyllactosamine $[\text{GlcNAc}\beta(1-3)\text{Gal}\beta(1-4)]_n$ structures are the preferred substrate. Sulfated structures such as keratan sulfate are also cleaved. Branching and/or fucosylation of the substrate may decrease or eliminate cleavage. Sulfation of C-6 on galactose will block cleavage. Oligosaccharides of the neo-lacto group are cleaved at greatly reduced rates depending on the deviation from the preferred substrate.

For example, $\text{Gal}\beta(1-3)\text{GlcNAc}\beta(1-3)\text{Gal}\beta(1-4)\text{Glc}$ is cleaved at 5×10^{-5} the rate of keratan sulfate (see ref.4). Specificity is similar to the *Escherichia freundii* enzyme, except that it is limited to cleaving N-acetyllactosamine extensions on tetraantennary structures of erythropoietin (see ref 5).

E-XBG01 Endo- β -Galactosidase

Specifications - Protocol

Specific Activity

One unit of endo- β -Galactosidase is defined as the amount that will liberate one μ mole of reducing sugar per minute at 37°C and pH 5.8 from bovine corneal keratan sulfate.

Purity

Endo- β -Galactosidase is tested for contaminating protease as follows: 10 μ g of denatured BSA is incubated for 24 hours at 37°C with 2 μ l of enzyme. SDS-PAGE analysis of the treated BSA shows no evidence of degradation.

The production strain of E. coli has been extensively tested and does not produce any detectable glycosidases.

Directions for use

For glycoproteins:

1. Add up to 100 μ g of glycoprotein to a tube.
2. Add 4 μ l 5X buffer and water to 19 μ l.
3. Add 1 μ l enzyme.
4. Incubate at 37°C for 2 hrs.

Procedure for oligosaccharides:

Same as above except incubate from several hours to several days depending on the substrate. Add bovine serum albumen to 2 mg/ml to stabilize the protein during extended incubations.

References

1. Scudder, P., Uemura, K., Doby, J., Fukuda, M.N. & Feizi, T. (1983) Isolation and characterization of an endo- β -galactosidase from *Bacteroides fragilis* Biochem. J. 213, 485-494.
2. Scudder, P., Hanfland, P.I., Uemura, K. & Feizi, T. (1984) Endo- β -galactosidases of *Bacteroides fragilis* and *Escherichia freundii* hydrolyze linear but not branched oligosaccharide domains of glycolipids of the neolacto series. J. Biol. Chem. 259, 6586-6592.
3. Scudder, P., Tang, P.W., Hounsell, E.F., Lawson, A.M., Mehmet, H. & Feizi, T. (1986) Isolation and characterization of sulfated oligosaccharides released from bovine corneal keratan sulphate by the action of endo- β -galactosidase. Eur. J. Biochem. 157, 365-373.
4. Murata, T., Hattori, T., Amarume, S., Koicki, A. & Usui, T. (2003) Kinetic studies on endo- β -galactosidase by a novel colorimetric assay and synthesis of N-acetylglucosamine-repeating oligosaccharide β -glycosides using its transglycosylation activity. Eur. J. Biochem 270, 3709-3719.
5. Hokke, C.H., Bergwerff, A.A., Van Dedem, D.W., Kamerling, J.P., and Vliegthart, J.F. (1995) Structural analysis of the N- and O-linked carbohydrate chains of recombinant human erythropoietin expressed in Chinese hamster ovary cells. Sialylation patterns and branch location of dimeric N-acetylglucosamine units. Eur. J. Biochem. 228, 981-1008.

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